

DNA Sequencing Technologies

Aleksandra Radenovic

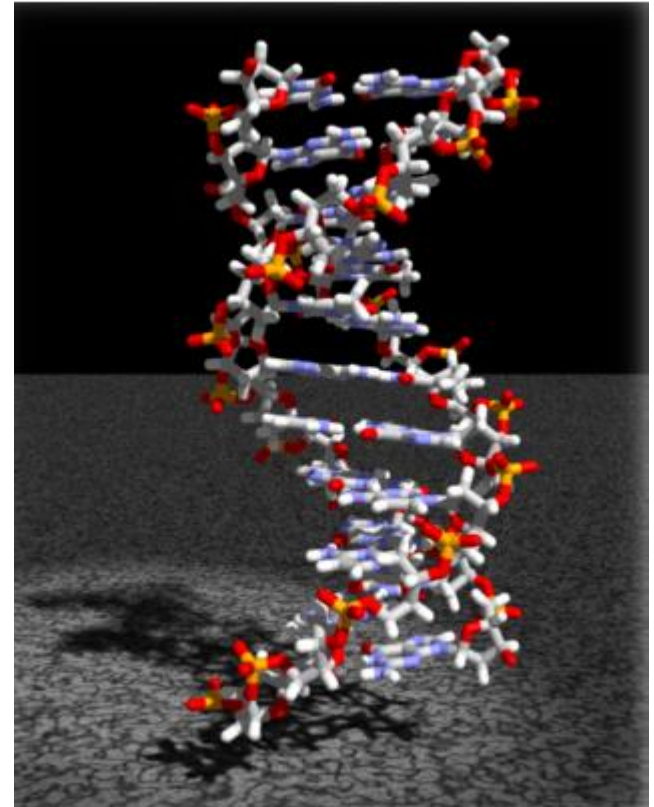
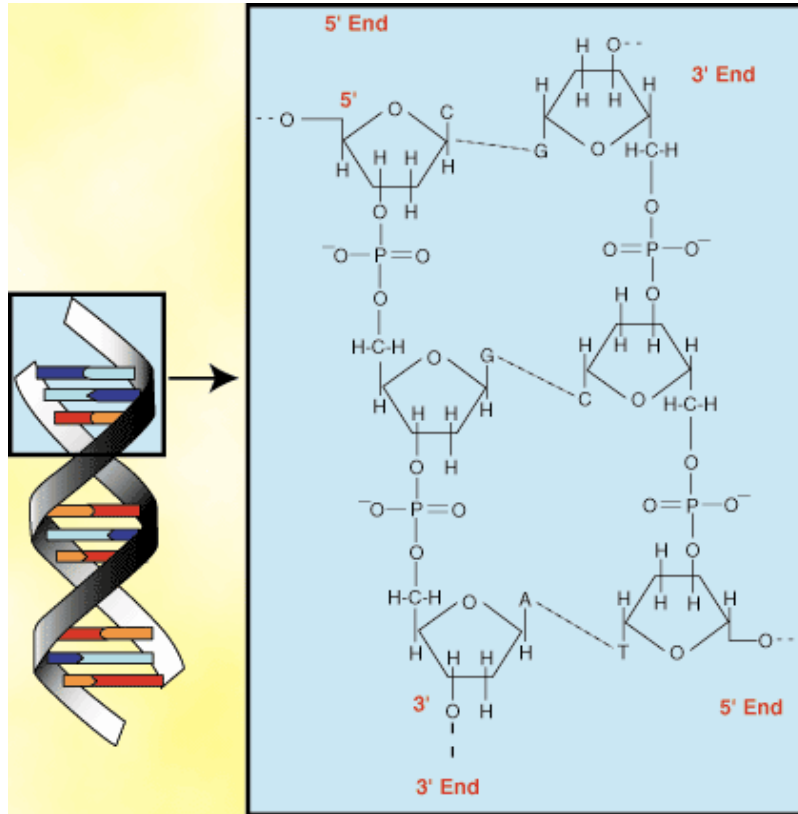
aleksandra.radenovic@epfl.ch

EPFL – Ecole Polytechnique Federale de Lausanne
Bioengineering Institute IBI



DNA –charged polymer

- DNA is a linear polymer molecule, i. e. a long chain of repeated subunits, called nucleotides. These come in four types, and thus DNA stores information as particular sequences of nucleotides



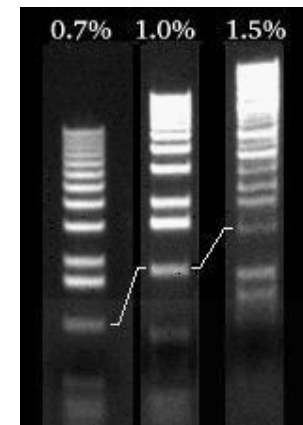
Phosphate carries negative charge

Gel electrophoresis

- Determining the length of a DNA molecule
- Gel electrophoresis involves driving charged molecules, such as DNA, through a porous gel matrix by means of an applied electric field. The gel matrix exerts a frictional force which increases with molecule size, and thus molecules of different
- sizes move at different speeds, separating as they move



$$f \cdot \vec{v} = q \cdot \vec{E}$$
$$\mu = \frac{\vec{v}}{\vec{E}} = \frac{q}{f} = \frac{z \cdot e}{f}$$



DNA Sequencing

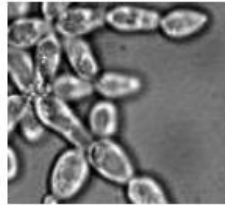
- Genome Projects



M. Tuberculosis



H. pylori



S. cerevisiae



C. elegans



Grape wine



Barley
Barley



Arabidopsis



Wheat



Medicago



Potato



Wasp



Maccaca



Neanderthal



James Watson



Cancer



Melon



Salamander



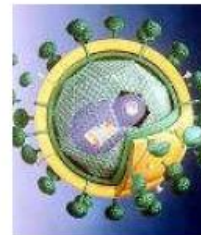
Bonobo



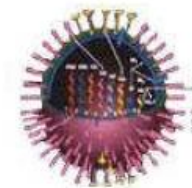
Honeybee



Mammut



HIV



H5N1



Toma


Why is Genome Sequencing Important?

- To understand how the genome as a whole works – how genes work together to direct growth, development and maintenance of an entire organism
- Understand how gene expression is regulated in a particular environment
- To study gene expression in a specific tissue, organ or tumor
- To study human variation
- To study how humans relate to other organisms
- To find correlations how genome information relates to development of cancer, susceptibility to certain diseases and drug metabolism (pharmacogenomics)

GOALS find sequence variability from cell to cell

DNA Sequencing

Israeli city to build dog poop DNA database

 September 19, 2008 by Mark O'Neill |

By **Mark O'Neill**

Contributing Writer, [GAS]

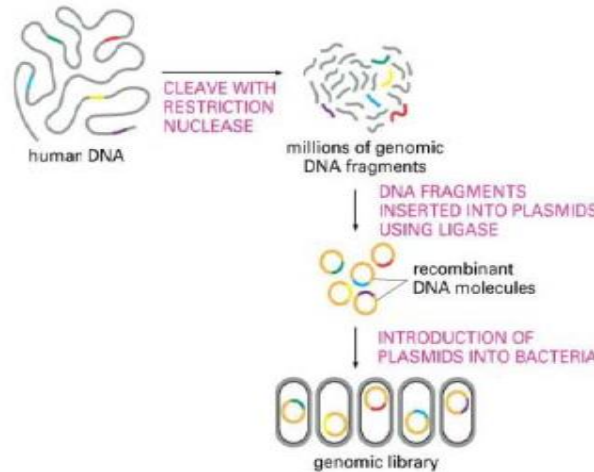
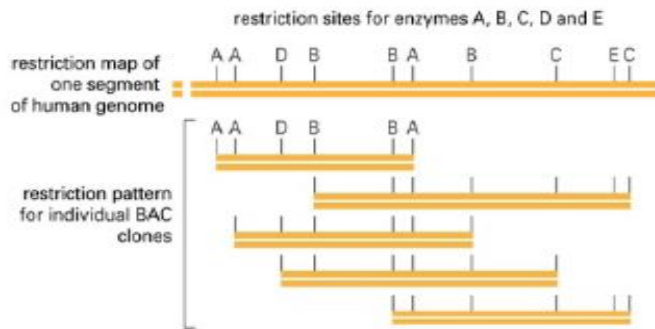


OK, now I *really* have heard everything!

The Israeli city of Petah Tikva is seemingly fed up with all the doggie poop lying around on their streets. So their solution is **to set up a dog poop DNA database**. To build this, all the dog owners of the city are being asked to take their canine pals into the local vet to have their dog's mouth swabbed. Rover or Tiddles can then have their DNA information entered into the computer.

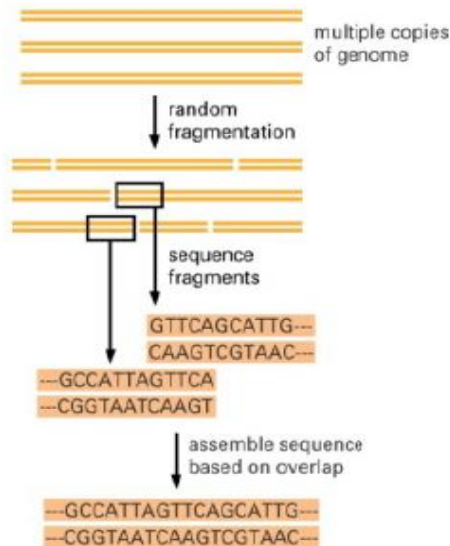
DNA Sequencing: How is Genome Sequencing Done?

- How is Genome Sequencing Done?
- *Clone by clone*



Create a crude physical map of the whole genome before sequencing with restriction enzymes. Break the genome into overlapping fragments and insert them into BACs and transfect into E.coli.

- *Shotgun sequencing*



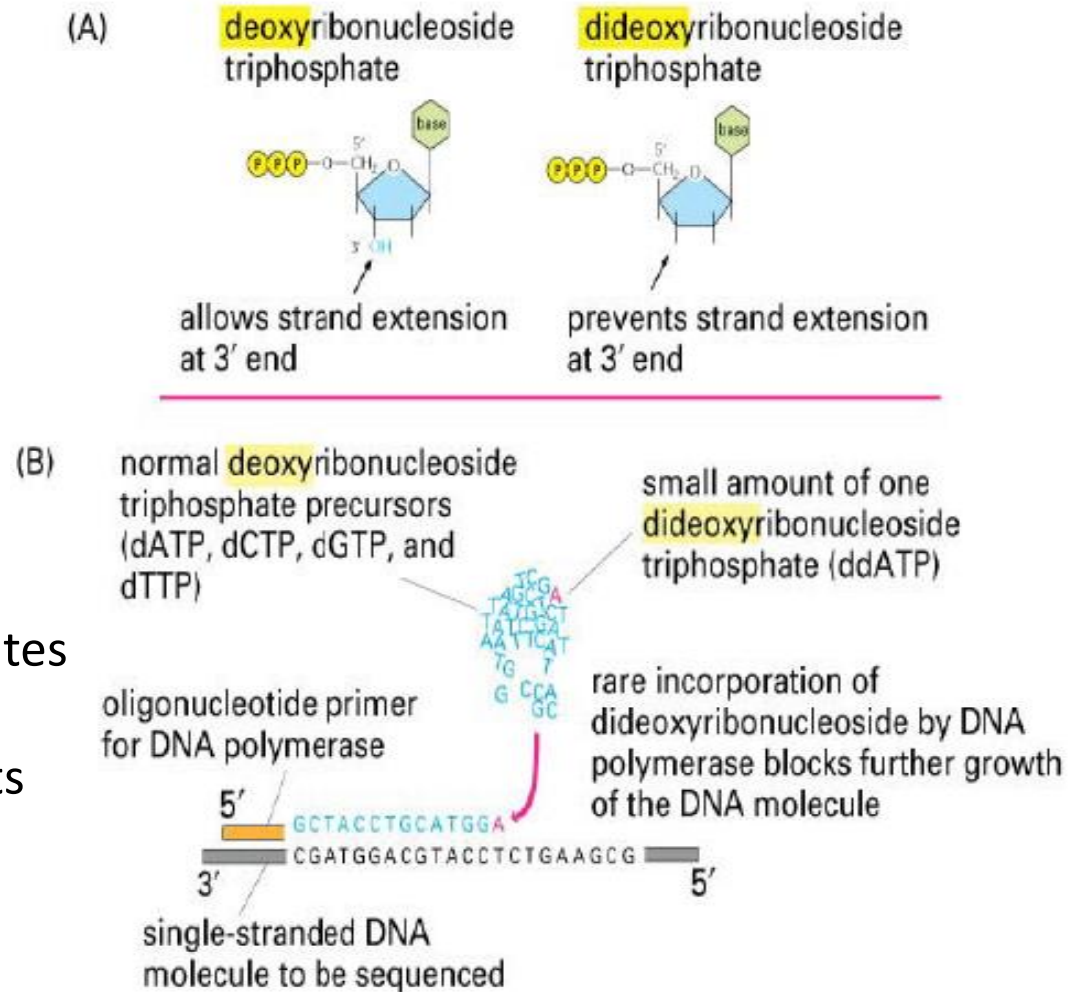
Break genome into random fragments, sequence each of the fragments and assemble fragments based on sequence overlaps.

DNA Sequencing: How is Genome Sequencing Done?

- **How is Genome Sequencing Done?**
- The key principle of the Sanger method was the use of [dideoxynucleotide](#) triphosphates (ddNTPs) as DNA chain terminators.

- “Normal” DNA synthesis:
 - DNA strand as template
 - Primer
 - Deoxynucleotides
 - Polymerase enzyme
 - Use several cycles to amplify

DNA synthesis is carried out in the presence of limiting amounts of dideoxy-ribonucleoside triphosphates that results in chain termination through chain termination fragments of distinct sizes are generated that can be separated by gel electrophoresis



DNA Sequencing: How is Genome Sequencing Done?

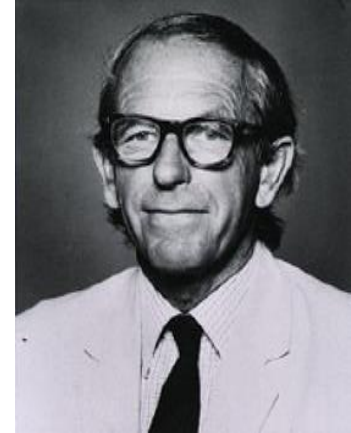
- Original method used radio-labeled primer or dideoxynucleotides This method required four separate DNA synthesis reactions to be separated by electrophoresis in four parallel lanes. The gel needs to be dried, exposed to film, developed and manually read. Approx. 150 bases read length

Summary – Sequencing Method Established

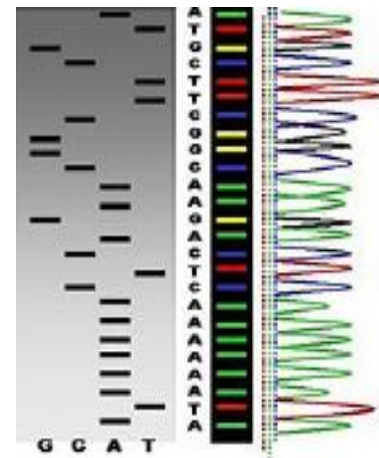
- Need of four reactions in parallel
- Heat labile polymerase
- Use of radioactivity
- Low resolution on gels
- Approx. 150 nucleotides read length
- time consuming

Improvements

- Use of fluorescently labeled dideoxynucleotides /one-lane electrophoresis
- Introduction of capillary electrophoresis to increase resolution (up to **1,000** ntes)
- Use of heat stable polymerase (Taq) Automation



Frederick Sanger
Nobel Prize (1980)



DNA Sequencing: How is Genome Sequencing Done?

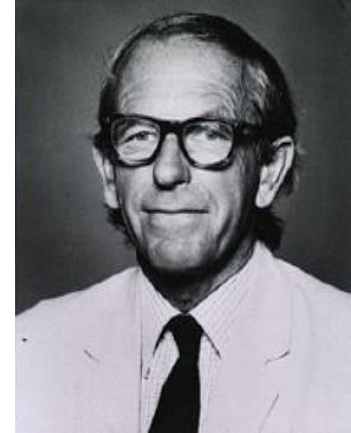
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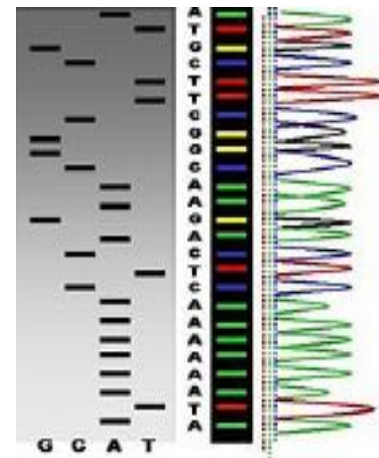
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Frederick Sanger
Nobel Prize (1980)



Human Genome Project Sequencing Centers



Whole Genome Shotgun Sequencing



Personalized genome Sequencing

Goals:

Link genome with phenotype
Provide personalized diet and medicine

(???) designer babies, big-brother insurance companies

Timeline:

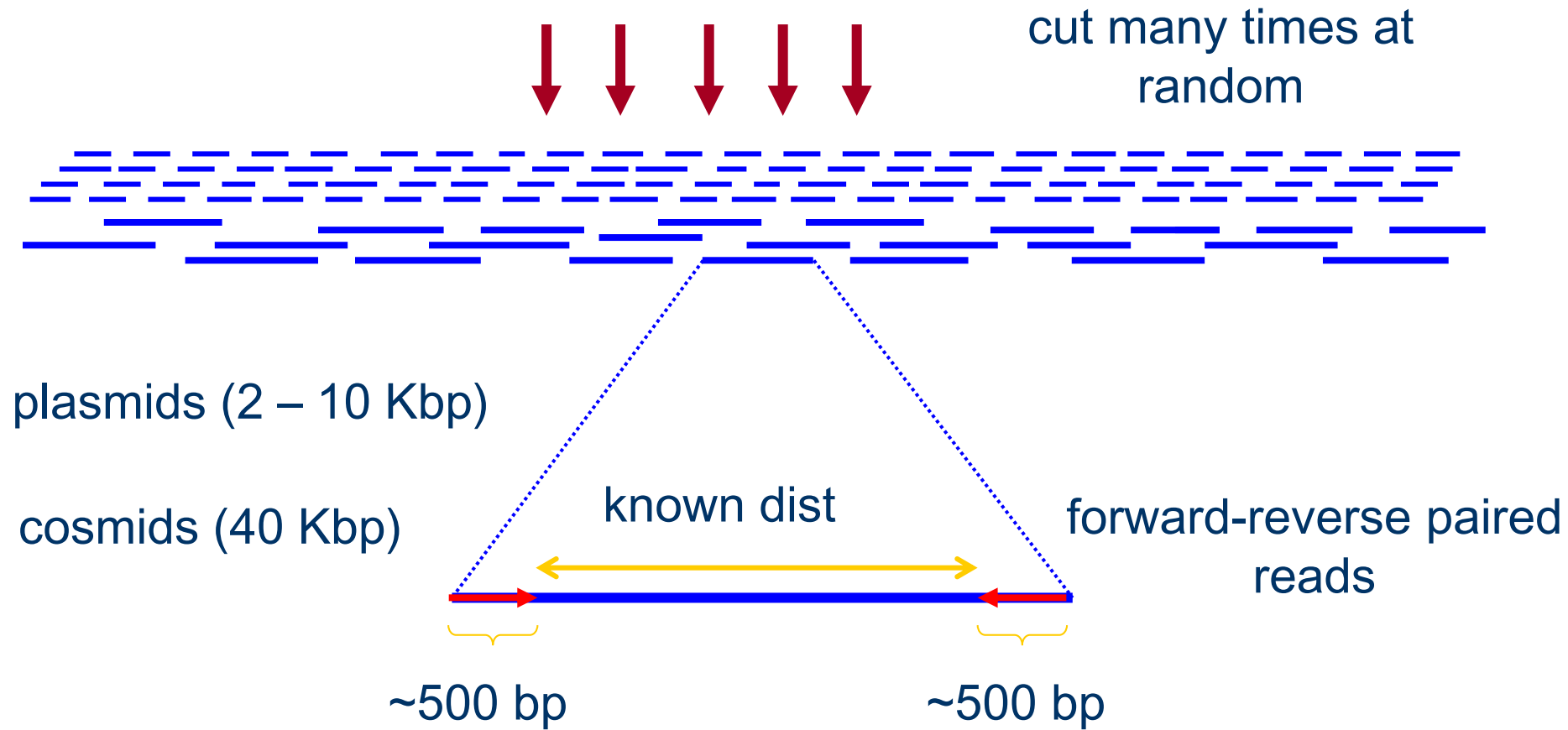
Inexpensive sequencing: 2010-2017

Genotype–phenotype

Personalized drugs: 2017-???

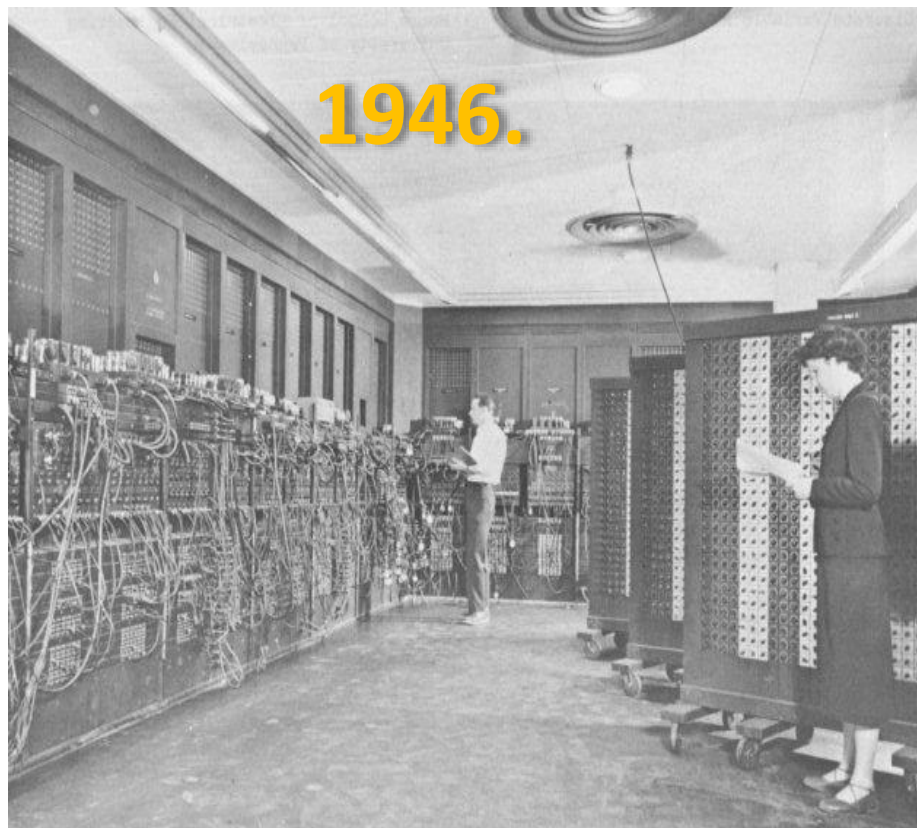
Whole Genome Shotgun Sequencing

Genome



Successful technology

Affordable, reliable, straightforward to use, and easy to adapt to new applications

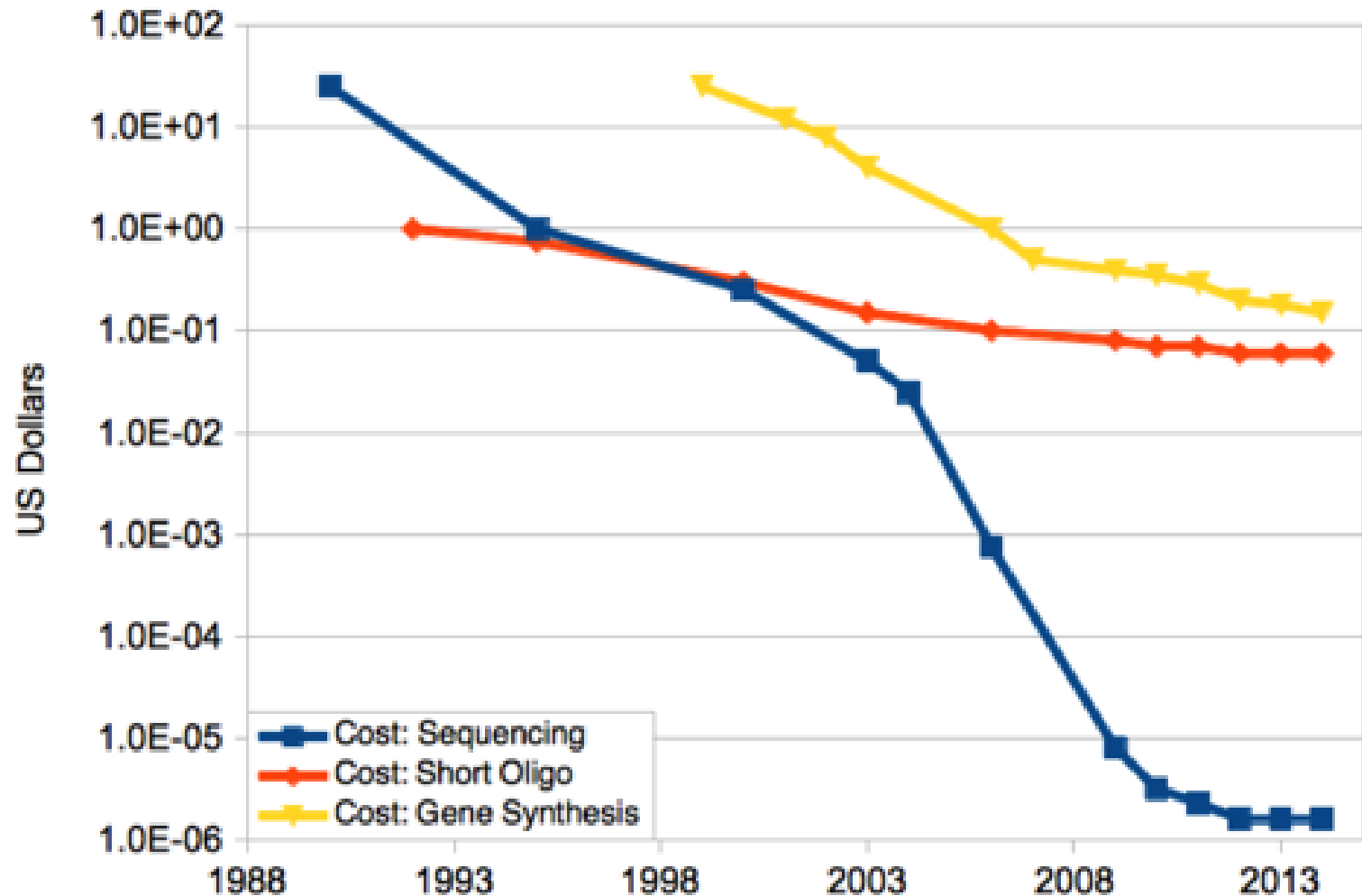


<http://en.wikipedia.org/wiki/ENIAC>



http://en.wikipedia.org/wiki/DNA_sequencer

Price per base for DNA sequencing and synthesis



Data from Rob Carlson www.synthesis.cc

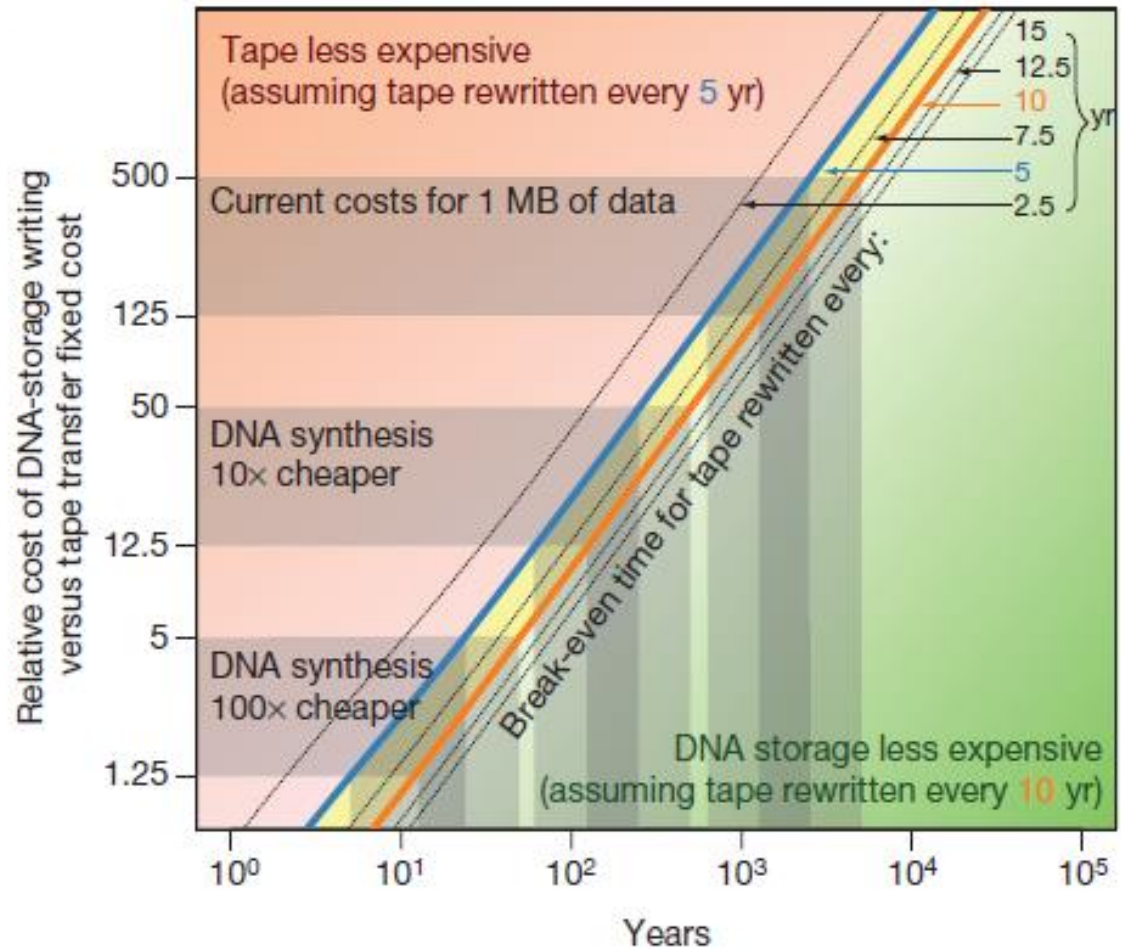
Year

Synthesized DNA- for information storage

effective solution for rarely accessed archives

high-capacity and low-maintenance

The speed of DNA-storage
writing and reading are not competitive with current technology

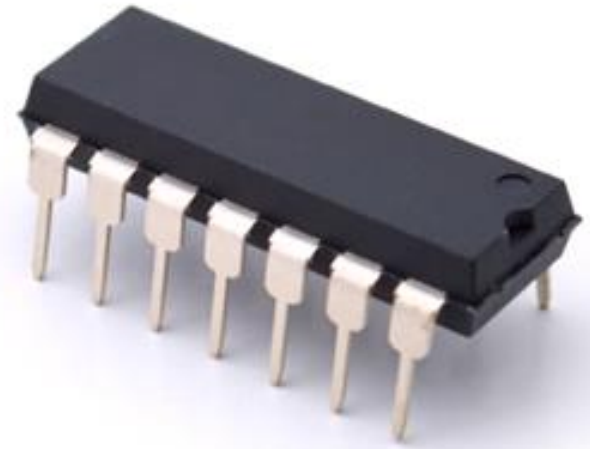


Could we advance DNA reading and writing?



Vacuum tubes: slow, expensive, fragile

Transistors: much simpler, much smaller, much cheaper, more reliable, no warm up, much faster.



Integrated circuits: miniaturization added to all the existing benefits, enabled unthought-of possibilities

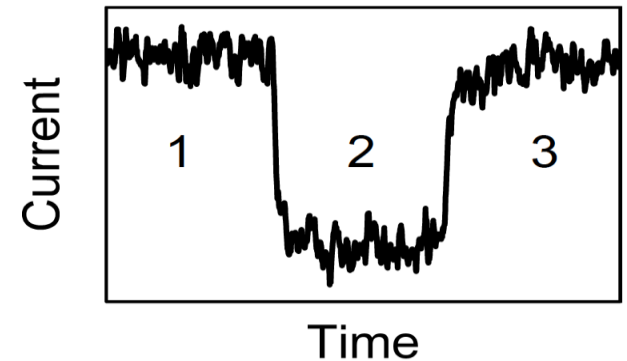
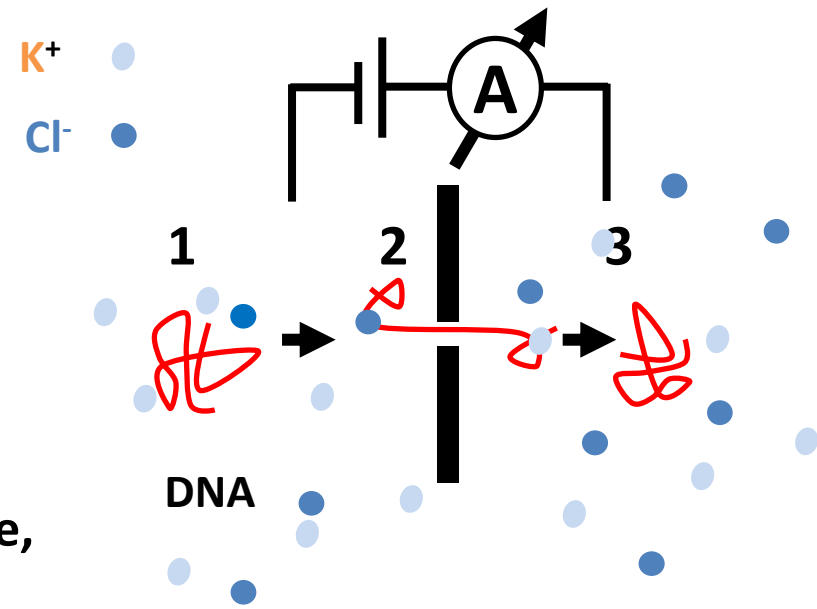
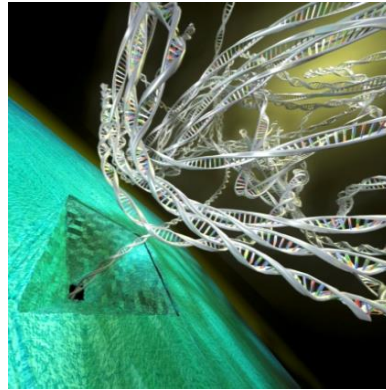
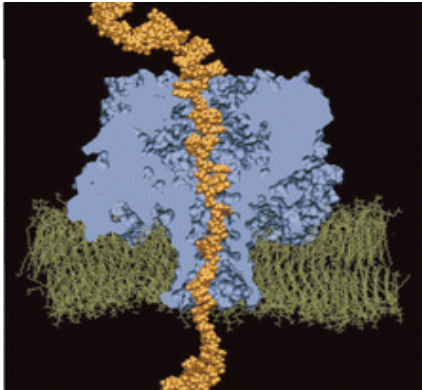
Nanopores

Single molecules –sensors

Sensitive to atomic composition

Sensing is intrinsic, no bleaching, nondestructive, repeatable

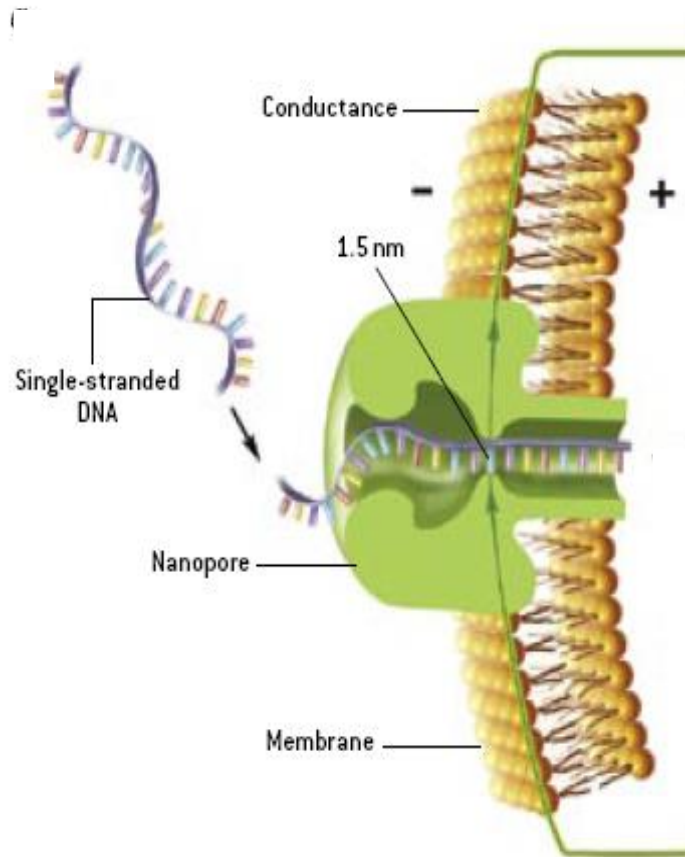
Sensing occurs in solution



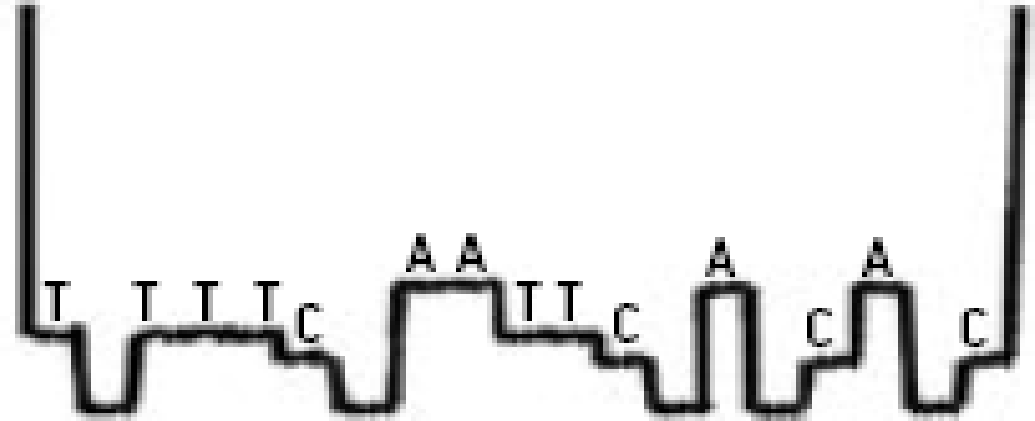
$$I = \frac{U}{R} = UG$$

Akeson M. et al. **1999** Meller A. et al. **2000** Howorka S. et al. **2001**
Li J, et al. **2001**. Dekker C. et. al. **2007**

The Goal: Automated Rapid DNA Sequencing with Nanopores



Hypothetical readout



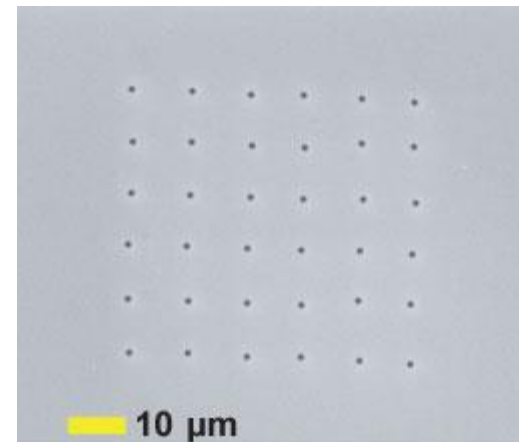
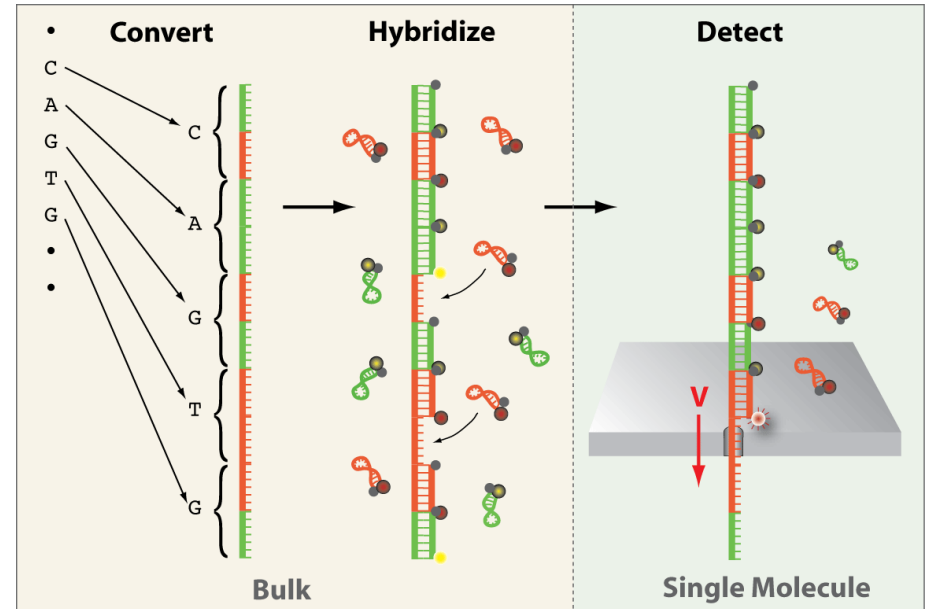
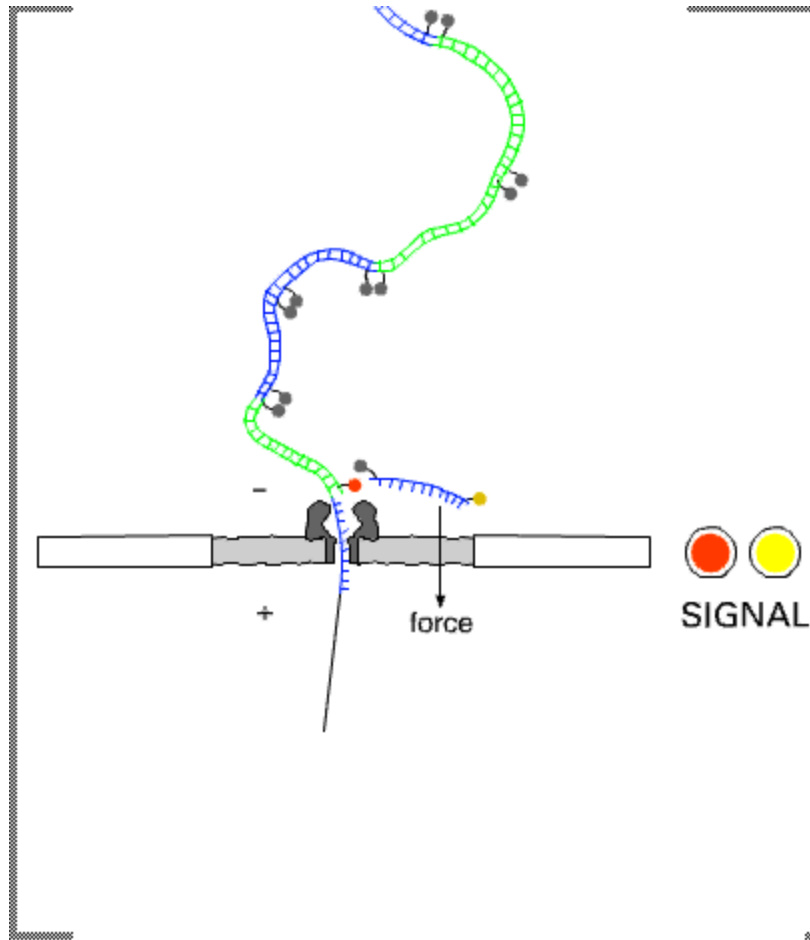
Advantages of nanopore sequencing:

- long-read lengths
- single molecule
- no amplification
- label-free
- electrical detection

DNA can be sequenced using nanopores only if its dynamics through the pore can be controlled...

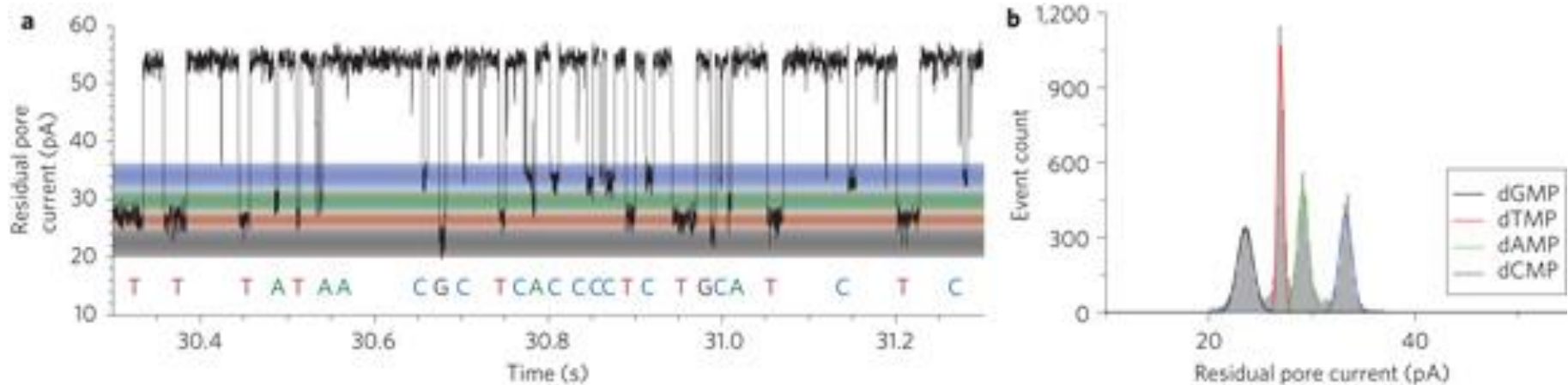
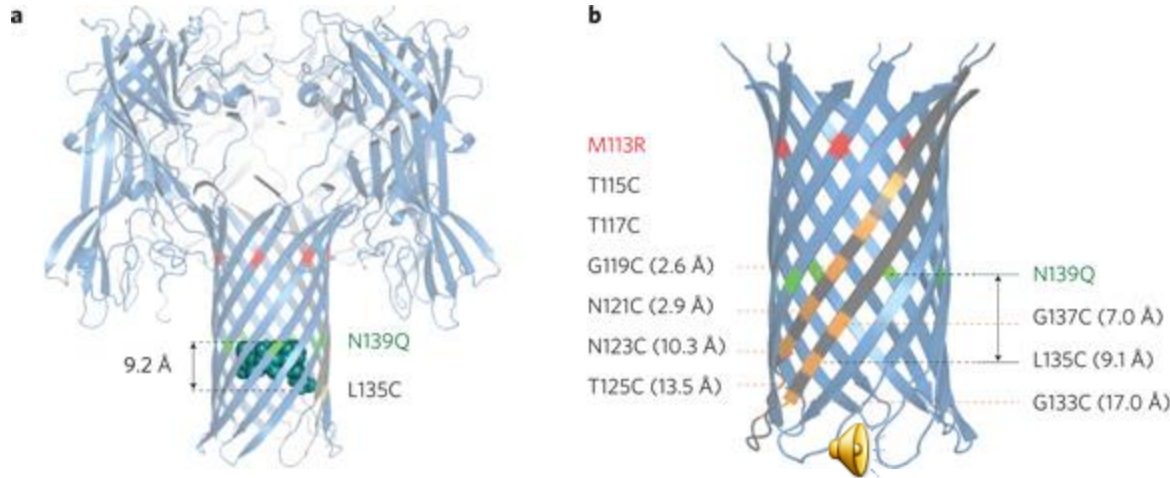
The Goal: Automated Rapid DNA Sequencing with Nanopores

- Ultra Fast DNA Sequencing Using Nanopores and Optical Probes



The Goal: Automated Rapid DNA Sequencing with Nanopores

- cyclodextrin covalently attached inside alpha hemolysin pore



Single-channel recording showing dGMP, dTMP, dAMP and dCMP discrimination, with coloured bands

Today: Oxford Nanopore



DNA

LETTER

doi:10.1038/nature16996

Real-time, portable genome sequencing for Ebola surveillance

Joshua Quick^{1*}, Nicholas J. Loman^{1*}, Sophie Duraffour^{2,3*}, Jared T. Simpson^{4,5*}, Ettore Severi^{6*}, Lauren Cowley^{7*}, Joseph Akoi Bore², Raymond Koundouno², Gytis Dudas⁸, Amy Mikhail⁷, Nobila Ouédraogo⁹, Babak Afrough^{2,10}, Amadou Bah^{2,11}, Jonathan H. J. Baum^{2,3}, Beate Becker-Ziaja^{2,3}, Jan Peter Boettcher^{2,12}, Mar Cabeza-Cabrerizo^{2,3}, Álvaro Camino-Sánchez², Lisa L. Carter^{2,13}, Juliane Doerrbecker^{2,3}, Theresa Enkirch^{2,14}, Isabel García-Dorival^{2,15}, Nicole Hetzelt^{2,12}, Julia Hinzmann^{2,12}, Tobias Holm^{2,3}, Liana Eleni Kafetzopoulou^{2,16}, Michel Koropogui^{2,17}, Abigael Kosgey^{2,18}, Eeva Kuisma^{2,10}, Christopher H. Logue^{2,10}, Antonio Mazzarelli^{2,19}, Sarah Meisel^{2,3}, Marc Mertens^{2,20}, Janine Michel^{2,12}, Didier Ngabo^{2,10}, Katja Nitzsche^{2,3}, Elisa Pallasch^{2,3}, Livia Victoria Patrono^{2,3}, Jasmine Portmann^{2,21}, Johanna Gabriella Repits^{2,22}, Natasha Y. Rickett^{2,15,23}, Andreas Sachse^{2,12}, Katrin Singethan^{2,24}, Inês Vitoriano^{2,10}, Rahel L. Yemanaberhan^{2,3}, Elsa G. Zekeng^{2,15,23}, Trina Racine²⁵, Alexander Bello²⁵, Amadou Alpha Sall²⁶, Ousmane Faye²⁶, Oumar Faye²⁶,



LETTER

doi:10.1038/nature16996

Real-time, portable genome sequencing for Ebola surveillance

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Viruses > ssRNA viruses > ssRNA negative-strand viruses > Mononegavirales > Filoviridae >

Ebolavirus - 5 complete genomes

Retrieve sequences:

-- Select

* The list view for each taxonomy node shows only the ne

* Unclassified/unassigned names are written in copper

Sort the genomes list by:

taxonomy

 Filter by host:

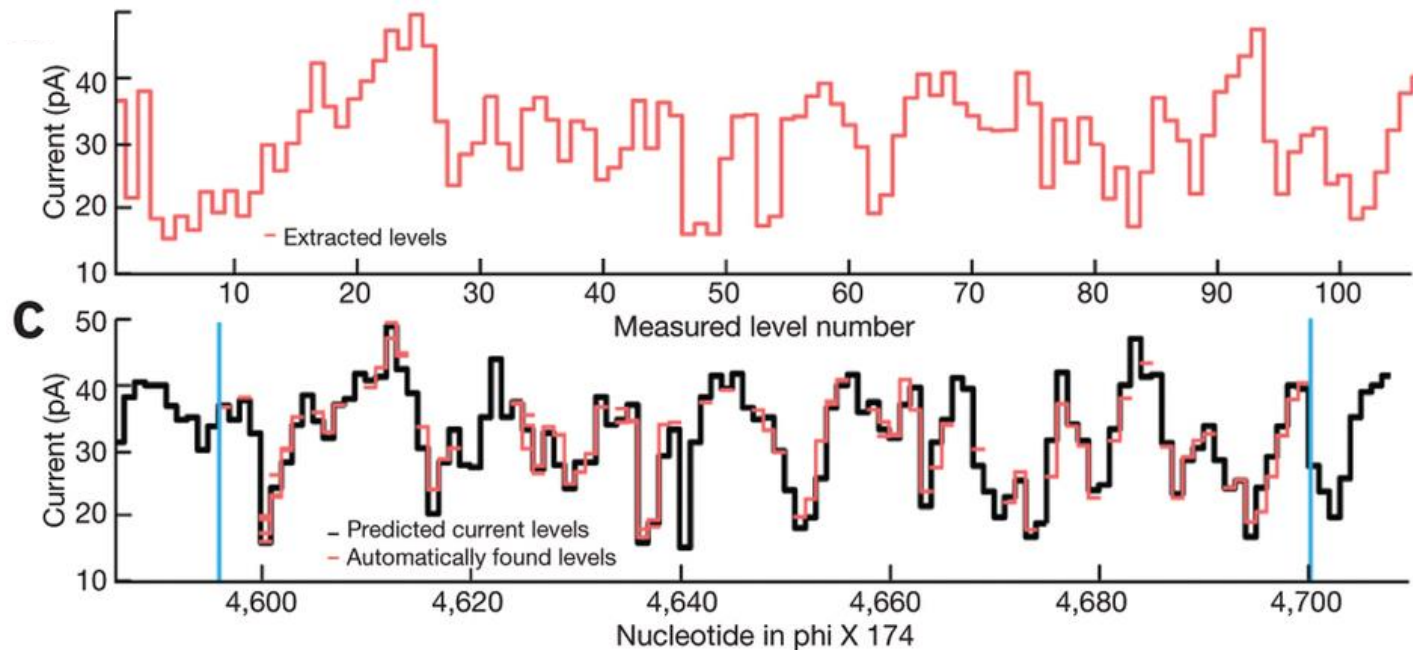
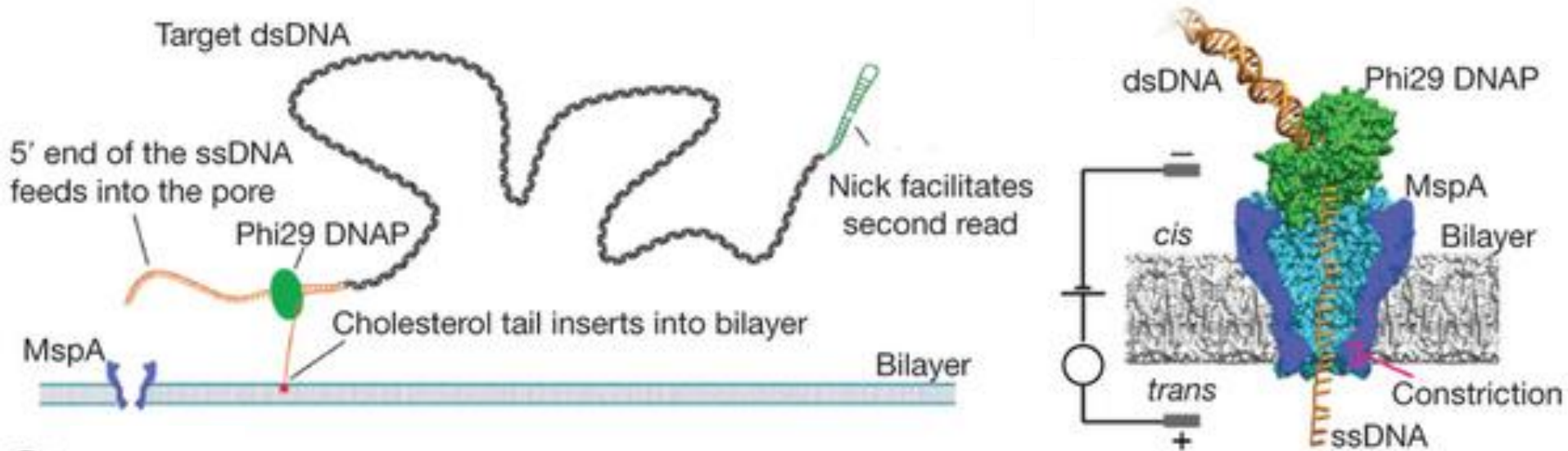
all hosts

Genome	Accession	Source information	Segm	Length	Protein	Neighbors	Host
Bundibugyo virus	NC_014373	isolate:Bundibugyo virus/H.sapiens-tc/UGA/2007/Butalya-811250	-	18940 nt	9	6	vertebrates, human
Reston ebolavirus	NC_004161	isolate:Reston virus/M.fascicularis-tc/USA/1989/Philippines89-Pennsylvania	-	18891 nt	8	7	vertebrates, human
Sudan ebolavirus	NC_006432	isolate:Sudan virus/H.sapiens-tc/UGA/2000/Gulu-808892	-	18875 nt	8	12	vertebrates, human
Tai Forest ebolavirus	NC_014372	isolate:Tai Forest virus/H.sapiens-tc/CIV/1994/Pauleoula-CI	-	18935 nt	9	1	vertebrates, human
Zaire ebolavirus	NC_002549	isolate:Ebola virus/H.sapiens-tc/COD/1976/Yambuku-Mayinga	-	18959 nt	9	1103	vertebrates, human

Real-time, portable genome sequencing for Ebola surveillance.



Nanopore strand sequencing











Human genome

ARTICLES

nature
biotechnology

OPEN

Nanopore sequencing and assembly of a human genome with ultra-long reads

Miten Jain^{1,13}, Sergey Koren^{2,13}, Karen H Miga^{1,13}, Josh Quick^{3,13}, Arthur C Rand^{1,13}, Thomas A Sasani^{4,5,13}, John R Tyson^{6,13}, Andrew D Beggs⁷, Alexander T Dilthey², Ian T Fiddes¹, Sunir Malla⁸, Hannah Marriott⁸, Tom Nieto⁷, Justin O'Grady⁹, Hugh E Olsen¹, Brent S Pedersen^{4,5}, Arang Rhie², Hollian Richardson⁹, Aaron R Quinlan^{4,5,10}, Terrance P Snutch⁶, Louise Tee⁷, Benedict Paten¹, Adam M Phillippy², Jared T Simpson^{11,12}, Nicholas J Loman³ & Matthew Loose⁸

We report the sequencing and assembly of a reference genome for the human GM12878 Utah/Ceph cell line using the MinION (Oxford Nanopore Technologies) nanopore sequencer. 91.2 Gb of sequence data, representing ~30x theoretical coverage, were produced. Reference-based alignment enabled detection of large structural variants and epigenetic modifications. *De novo* assembly of nanopore reads alone yielded a contiguous assembly (NG50 ~3 Mb). We developed a protocol to generate ultra-long reads (N50 > 100 kb, read lengths up to 882 kb). Incorporating an additional 5x coverage of these ultra-long reads more than doubled the assembly contiguity (NG50 ~6.4 Mb). The final assembled genome was 2,867 million bases in size, covering 85.8% of the reference. Assembly accuracy, after incorporating complementary short-read sequencing data, exceeded 99.8%. Ultra-long reads enabled assembly and phasing of the 4-Mb major histocompatibility complex (MHC) locus in its entirety, measurement of telomere repeat length, and closure of gaps in the reference human genome assembly GRCh38.

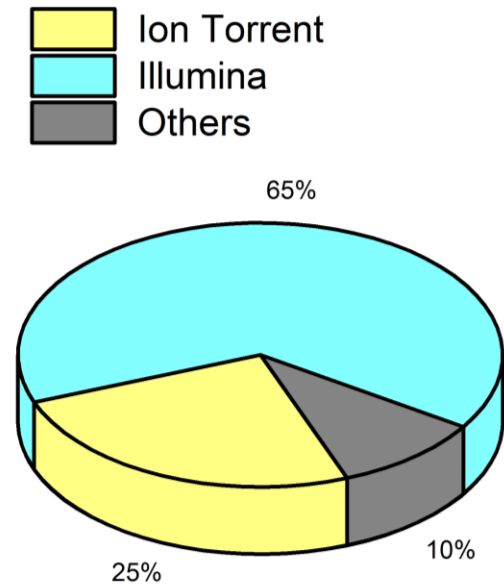
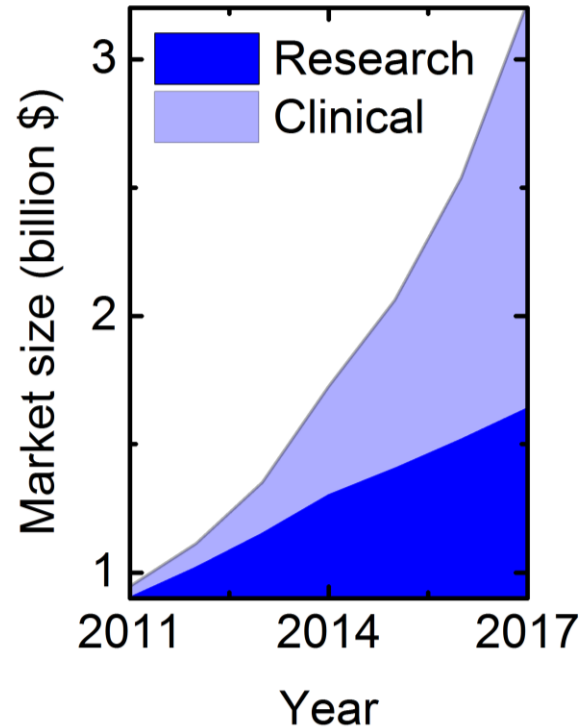
Sequencing applications

Personalized medicine- drug development

Cancer genomics

Microbial genetics – (Human microbiome)

Population genetics – evolution



Emerging nanopore applications beyond DNA sequencing

Protein Biomarker Analysis

Characterization, identification, and counting of individual protein molecules or nanoparticles

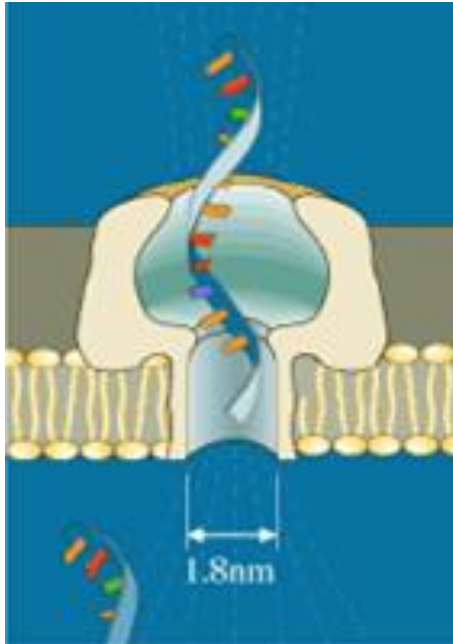
Peptide Sequencing

Water Desalination

Power Generation

Why Synthetic Pores? Disadvantages of protein pores

α hemolysin



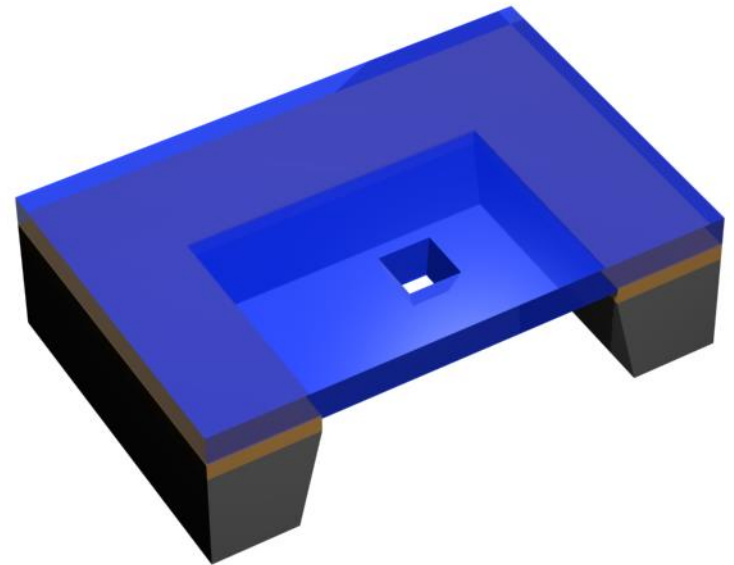
Fixed diameter

Typical velocity:
 $\sim 1 \text{ base}/\mu\text{s} = 0.3 \text{ mm/s}$

Short life time

Some protein pore display self gaiting

Synthetic



Tunable diameter

Typical velocity:
 $\sim 3 \text{ base}/\mu\text{s} = 10 \text{ mm/s}$

Usable up to several days

Why Synthetic Pores?

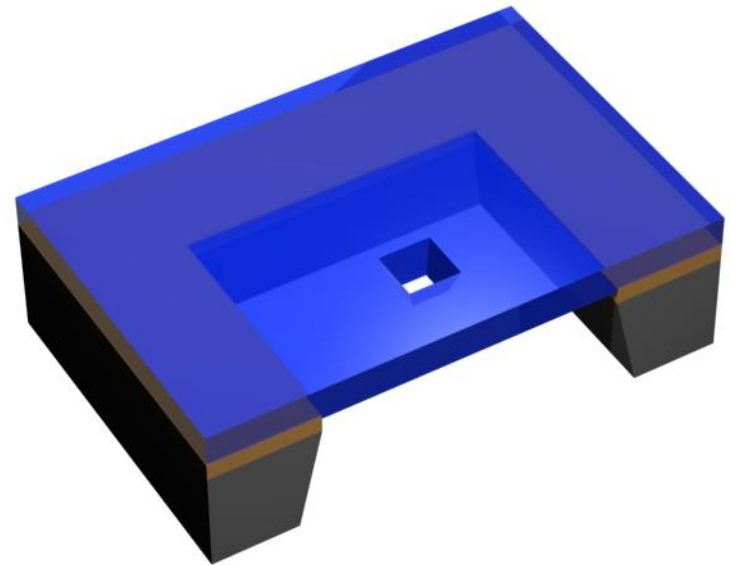
precise control over the location and chemical properties of the pores (pH, solvents, ionic strengths, oxidizers, etc.)

sufficiently **stiff** to permit high resolution distance detection and application of forces > 60 pN to the polymer (Physically robust (vibrations, pressure changes)

directly **compatible** with numerous detection schemes, including optical trapping, pA current measurements, and solid state detectors deposited near a pore

Tunable size and interface ,Fixed coordinates (pore position always the same)

Synthetic



Tunable diameter

Typical velocity:
 ~ 3 base/ μs = 10 mm/s

Usable up to several days

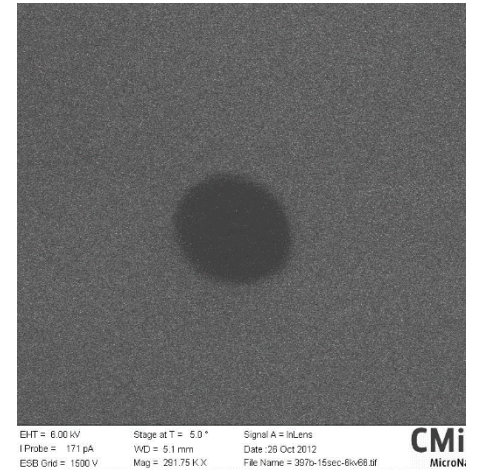
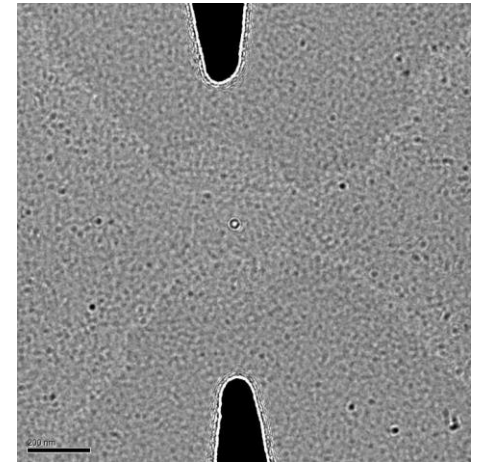
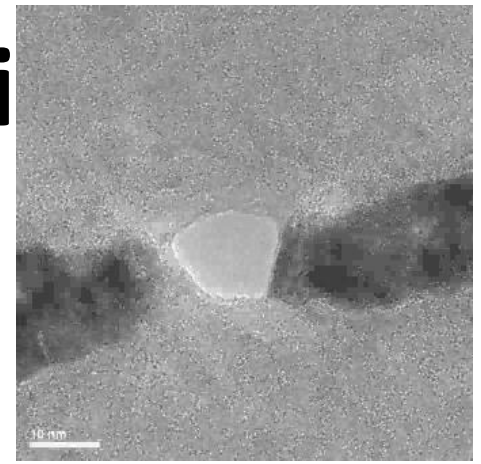
Nanopores - materi

Nanopore material – dictates application

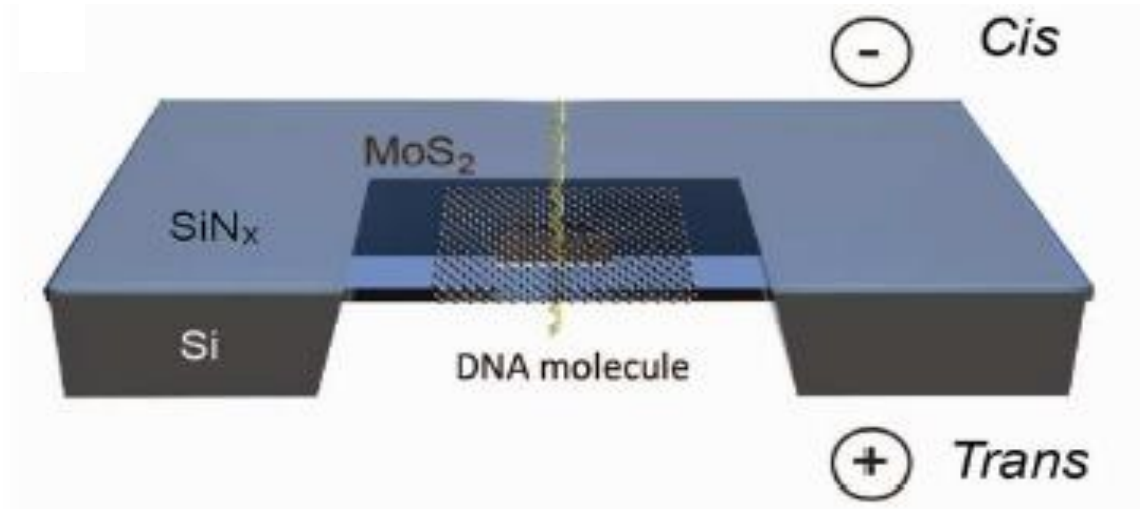
Silicon nitride nanopores – gold standard for solid state nanopores **5-20 nm thick**

2D material nanopores: graphene and molybdenum disulfide (MoS_2)-**0.3-0.7 nm thick**

Glass/Quartz nanocapillaries



Nanopore sensitivity – 2D materials



- **Sensitivity** and **Selectivity**: Transverse current, monolayer (Graphene, molybdenum disulfide MoS₂ other 2D materials or very thin nitride)
- **Statistics**: high throughput via multiplexing using concomitant detection with- for example integrated FETs
- **Speed** (time resolution) slowing down ultra fast translocation
- **Price** – low requires fabrication without TEM

Garaj S, et al. **2010** Nature, Merchant CA, et al. **2010** Nano Lett. Schneider GF, et al. **2010**. Nano Lett Liu, Radenovic et al. **2014** ACS Nano. Traversi ,Radenovic et al. Nature Nano. **2013**.

Research Groups

- D. Brantob, Jene Golovchenko , Harvard nanopore Group
- Dave Deamer, University of California at Santa Cruz
- Cees Dekker, Delft University of Technology
- Jiali Li, University of Arkansas
- Andre Marziali, University of British Columbia
- Amit Meller Laboratory, Boston University
- Gregory Timp, Nano-Bio Group, University of Illinois, Urbana-Champaign
- Aleksei Aksimentiev, UIUC
 - Sequencing a DNA molecule using silicon integrated circuit nanopores.
- Rahid Bashir, Samir Iqbal ,Perdue University
- Jacob Schmidt , Bioengineering, UCLA